



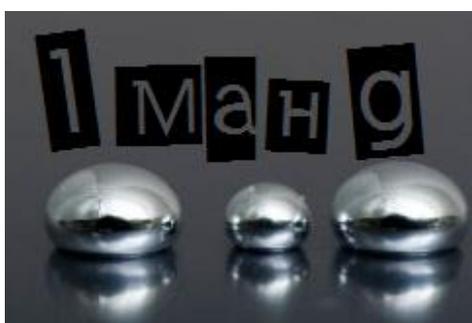
CLÉAIRE



## SNOWMAN NETWORK

Knowledge for sustainable soils

Project No. SN-03/08



Enhanced knowledge in mercury fate and transport  
for Improved Management of Hg soil contamination

# Literature Review on Mercury Speciation Soil Systems Under Oxidizing Conditions

Start date of project: 01.10.2011

Project duration: 27 months (2011-2013)

End date of project: 31.12.2013

Date of report: 06.08.2013

WP coordinator: Diederik Jacques (SCK•CEN)

Redactors: Bertrand Leterme (SCK•CEN), Diederik Jacques (SCK•CEN)

Revision: Vf





## Table of content

Summary .....	1
1 Mercury in the soil.....	3
1.1 Mercury cycle and transport .....	3
1.2 Sites of anthropogenic contamination .....	4
2 Aqueous speciation of mercury .....	7
2.1 Mercury oxidation and reduction .....	7
2.2 Mercury methylation and demethylation.....	8
2.3 Inorganic complex forms .....	9
2.4 Organic complex forms.....	9
3 Phase transitions of mercury .....	12
3.1 Dissolution .....	12
3.1.1 Dissolution of HgS(s) and HgO(s) .....	12
3.1.2 Dissolution from non-aqueous liquid form .....	12
3.2 Sorption .....	13
3.2.1 Types of Sorbent .....	13
3.2.2 Effect of dissolved species ( $S^{2-}$ , $Cl^{-}$ and DOC), pH and heat .....	14
3.2.3 Equilibrium or kinetic sorption, hysteresis .....	14
3.3 Volatilization.....	15
4 Conclusion .....	17
5 References .....	18
6 Annex.....	25
6.1 NAPL migration and entrapment .....	25

## List of figures

Figure 1 – Mercury sources (in blue), sinks (in orange) and phases in soil (in grey).....	3
Figure 2 – Dominant Hg aqueous species under oxic conditions (a) in the absence and (b) in the presence of 10 nM of low molecular mass thiols (RS), and (c) for MeHg under oxidized conditions in the presence of DOC 50 mg/L (having 2.3 µM of thiol functional groups, DOM-RS) and (d) a pH of 7. Marine conditions are indicated by black areas, and freshwater conditions by gray areas. Source : Skyllberg (2012). .....	10
Figure 3 – Dominance diagrams for soils and sediments with 100 ng Hg/g dry mass and 90 mass% pore water content. (a) The solid phase contains 15% organic C (NOM having a concentration of thiol functional groups corresponding to 0.8 mM on a pore water basis): the concentration of inorganic sulfur is 0.1 mM. Solid lines show the stability of mercury species, black broken lines show the stability of water and grey broken lines show the stability of sulfur species. (b) Limiting concentrations of S(-II)(= [H <sub>2</sub> S] + [HS <sup>-</sup> ]) for the precipitation of metacinnabar in sediments with different concentrations of organic C. Regulation of S(-II) by the solubility of FeSm is indicated at freshwater (area bordered by dotted line) and marine conditions (black field). The stability of FeSm is in equilibrium with Fe <sup>2+</sup> concentrations of 10 <sup>-3</sup> to 5×10 <sup>-5</sup> M in freshwater and 10 <sup>-6</sup> to 5×10 <sup>-8</sup> in marine environments. Source : Skyllberg (2012). .....	11
Figure 4 – Hg NAPL saturation profile at different time steps, for an illustrative simulation with parameters given in Table 1. ....	28

## List of table

Table 1 – Parameters of the HSSM model for simulating a test case of Hg NAPL migration in the vadose zone. ....	27
---	----

# Summary

This report consists of a literature review of factors and processes affecting mercury (Hg) fate in soils. Considering Hg cycle from the perspective of soil systems, Hg sources include atmospheric wet and dry deposition, geogenic sources and anthropogenic contamination. The main Hg sinks include volatilization to the atmosphere, and transport to groundwater or surface water. Plant root uptake from soils is extremely low and in general can be neglected.

Mercury can exist in three valence states:  $\text{Hg}^0$ ,  $\text{Hg}^{\text{I}}$  and  $\text{Hg}^{\text{II}}$ . However, the presence of  $\text{Hg}^{\text{I}}$  in soils is negligible compared to  $\text{Hg}^0$  and  $\text{Hg}^{\text{II}}$ , and the latter is dominant. The main dissolved Hg species in soil systems is  $\text{Hg}^{\text{II}}$  complexes with various inorganic and organic ligands. The key factors controlling Hg aqueous speciation are pH, ionic strength and redox potential. Concentrations of dissolved organic matter (DOM), dissolved oxygen, sulfide and suspended solids in solution can also play a role.

In oxidized soil conditions the predominant inorganic mercury species are  $\text{Hg}(\text{OH})_2$ ,  $\text{HgCl}_2$ ,  $\text{HgOH}^+$ ,  $\text{HgS}$ , and  $\text{Hg}^0$ . The first two are the most frequent. (Mono)methylmercury (MMHg) and dimethylmercury (DMHg) are the methylated forms of Hg that can be synthesized in soil or aqueous systems (by microbial activity). Generally, the concentration of MMHg is less than 2% of the total mercury concentration in soil systems, while DMHg is nearly absent.

Strong interactions between mercury and organic matter are observed. Hg complexes with thiol groups and humic and fulvic acids are particularly important. Migration of Hg to deeper soil layers is often suggested to be most effective if Hg is bound to soluble organic complexes (i.e. colloid-facilitated transport).

Different phase transitions of mercury occur in soil systems. Solid  $\text{HgS}$  (cinnabar) is the common ore of mercury. In the presence of  $\text{Cl}^-$  or DOM,  $\text{HgS}(\text{s})$  dissolution is observed and precipitation is inhibited. In case of anthropogenic contamination, Hg can be present in the form of a (dense) non-aqueous phase liquid or (D)NAPL, which can further dissolve or volatilize in soil systems.

Similarly to what is observed for Hg aqueous complexation with organic ligands, sorption capability of  $\text{Hg}^{2+}$  is highest for humic and fulvic substances and for molecules containing a reactive thiol functional group. Adsorption rate depends mainly on initial  $\text{Hg}^{2+}$  concentration, organic matter content and ions in solution.

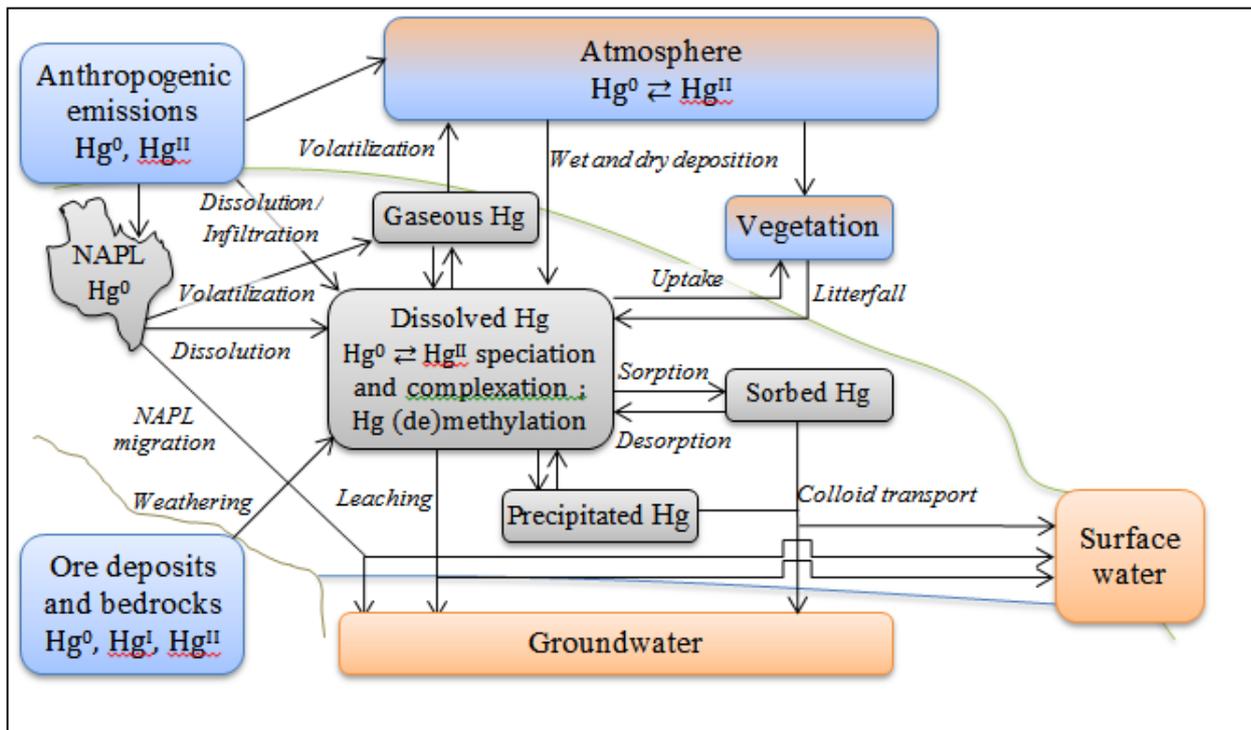
Volatilization of mercury occurs in soils and Hg vapor flux to the atmosphere can be important in case of anthropogenic contamination. Volatilization can be described as a three-step process, in which  $\text{Hg}^{\text{II}}$  is first reduced to  $\text{Hg}^0$ .  $\text{Hg}^0$  is then diffused to the soil surface and finally volatilized across the soil-air boundary layer into the atmosphere. Sunlight and heat are the main factors driving Hg volatilization.



# 1 Mercury in the soil

## 1.1 Mercury cycle and transport

Many reviews or descriptions of mercury fate and transport in the environment take a global perspective (e.g. UNEP, 2002) where soils are only a compartment (pool) of Hg. If not assessed from a global perspective, Hg cycle is often restricted to either atmospheric (e.g. Schroeder and Munthe, 1998) or aqueous (wetland, river, lake, ocean; e.g. Sunderland, 2009) systems. Soils are then often seen as a sink/source of mercury, but to date we found no study giving a comprehensive description of all sources/sinks and transport processes relevant for Hg fate in soils. The cycle of Hg fate and transport from the perspective of soils is depicted in Figure 1. The objective of this figure is to give a qualitative rather than a quantitative description of mercury fate in soils. It is important to note that Figure 1 does not show all components and fluxes of the global Hg cycle, but only the most relevant for the vadose zone. For example, volatilization from ore deposits to the atmosphere, erosion and runoff of soil particles to surface water, or mercury fate in aqueous systems (river, lake, oceans, sediments...) are not shown.



**Figure 1 – Mercury sources (in blue), sinks (in orange) and phases in soil (in grey).**

Mercury may be present under different phases in soil systems: (i) dissolved in the aqueous phase as a free ion ( $\text{Hg}^{2+}$ ) or complexed with inorganic and/or organic ligands, (ii) metallic (or elemental)  $\text{Hg}^0$  as a non-aqueous liquid phase (NAPL), (iii) sorbed on soil minerals and insoluble organic matter, (iv) in the gas phase, and (v) in solid (precipitated) phase. Aqueous speciation of mercury is reviewed in section 2 and phase transitions are discussed in section 3.

Sources of Hg in soils consist of (Figure 1): (i) atmospheric wet and dry deposition and litterfall, (ii) geogenic (or lithogenic) mercury, and (iii) anthropogenic contamination.

In the atmosphere, mercury in the form of gaseous  $\text{Hg}^0$  is dominant (>95%; Zhang et al., 2009). Reviews of atmospheric mercury cycle, chemistry and modeling can be found in (Lin and Pehkonen, 1999; Schroeder and Munthe, 1998; Subir et al., 2011). Mercury wet and dry deposition is mostly in the form of inorganic  $\text{Hg}^{\text{II}}$  (Zhang et al., 2009), possibly in the form of  $\text{HgCl}_2$ ; a large fraction being associated with particles (Schlüter, 2000). Deposition in the form of monomethylmercury ( $\text{CH}_3\text{Hg}^+$ ) is usually reported to be less than 5% of total deposition (Schlüter, 2000). From the perspective of soil systems, litterfall can be considered as a Hg source. Atmospheric Hg is the main source of Hg in aboveground vegetation tissues and when these fall to the ground Hg can be further released in soils (Obrist et al., 2011).

Geogenic (or lithogenic) sources provide Hg via oxidation and weathering of ore deposits and bedrocks. Mercury may then be transported to soils via  $\text{Hg}^0$  degassing or via  $\text{Hg}^{\text{I}}/\text{Hg}^{\text{II}}$  transport in rising groundwater. One usually makes the distinction between Hg enriched areas (global “mercuriferous belt”) and background areas (Schlüter, 2000). The former show high rates of mercury degassing and/or weathering. It is also in these areas that most of the mercury mines are found (mainly in the form of  $\text{HgS}$ (s) ore). More details on the location of Hg enriched areas and their origin can be found in (Gustin et al., 2006; Schlüter, 2000). Background areas are zones where the geogenic flux of mercury to soils and atmosphere is small or negligible. In the field, methods exist to discriminate between atmospheric and geogenic sources of soil Hg (Guedron et al., 2006).

Anthropogenic soil contamination can result from direct sources (spilling, mine tailing, landfills, polluted sewage sludge...) but it can also occur indirectly, for example via deposition of prior atmospheric emissions. Mason et al. (1994) estimated that about two thirds of atmospheric Hg is directly or indirectly from anthropogenic sources. Direct contamination of soils can occur in the form of a non-aqueous phase liquid (NAPL) phase, which can then percolate, volatilize and dissolve. Specific aspects of soil contamination from anthropogenic sources and case studies are presented in section 1.2.

Mercury sinks from the perspective of soil systems are (Figure 1): (i) atmosphere, (ii) plants (via root uptake), (iii) groundwater, and (iv) surface water. Irreversible sorption sites are sometimes considered as a sink in the budget of soil Hg, but are not treated as sinks in this report, because this process does not change the total Hg budget in the soil.

Volatilization to soil gaseous phase and eventually to the atmosphere is discussed in section 3.3.

Plant root uptake from soils is extremely low and in general can be neglected (Pant et al., 2010; Schuster, 1991). Roots perform as a significant adsorption site and are therefore a barrier for Hg transport to foliage (Grigal, 2003). Therefore transport with the plants to the above ground parts is limited. It should be noted that some exceptions exist (e.g. Pérez-Sanz et al., 2012). Also, mercury phytoremediation can benefit from genetic engineering (Ruiz and Daniell, 2009) or association with rhizobacteria (Sorkhoh, 2010). Non terrestrial plants like mangroves can show significant Hg root uptake (Ding et al., 2011).

In the unsaturated zone, mercury transport to surface water (i.e. via subsurface flow) and to groundwater occurs via convection, dispersion, diffusion and colloid-facilitated transport. On sites of heavy contamination, the migration and entrapment of NAPL needs also to be taken into account. In the vadose zone, gaseous diffusion of mercury also participates in determining Hg fate in soils. Retardation of Hg in soils is mainly due to adsorption processes, which are examined in section 3.2.

## 1.2 Sites of anthropogenic contamination

Anthropogenic mercury contamination may result in much higher Hg concentrations in soil systems than from other sources. This has important consequences for assessing the fate and transport of Hg in soils, by affecting for instance  $\text{Hg}^{\text{II}}$  speciation in soils. Hg binding is preferentially on S-containing

functional groups of organic ligands. These are always in excess relative to Hg concentration in uncontaminated soils (see section 2.4), while contaminated sites may show much lower S/Hg ratios. It has also been found that in soil samples with anthropogenic contamination, the ratio of organomercury species to total mercury is usually lower than in uncontaminated areas (Boszke et al., 2008).

The activities most frequently cited as causes of anthropogenic contamination of the environment are (UNEP, 2002): mercury mining, gold and silver mining ( $\text{Hg}^0$  is used to amalgamate the metal), manufacturing (chlor-alkali plants, manometer spill), wood preservation, cemeteries (release of mercury from dental amalgams).

Logically, the Hg species present in the soil highly depend on the type of industry or activity that led to the contamination. Bloom et al. (2003) illustrated the variety of Hg speciation found at contaminated sites (gold mine tailings, HgS mines,  $\text{Hg}^0$  spill, chlor-alkali plant,  $\text{Hg}(\text{NO}_3)_2$  contamination from hydrogen bomb production). They concluded that the source Hg compound was important but also the time (aging) since mercury contamination, because phase transformation or retention mechanisms can have relatively slow kinetics. For example, even after decades, elemental mercury discharged by manometer spills or as an amalgam for mining, may remain as  $\text{Hg}^0$  “globules” or NAPL in the soil (Davis et al., 1997).

In a study of soil and aquifer contamination by  $\text{HgCl}_2$  used for wood preservation, Bollen et al. (2008) found that most Hg in soil was Hg bound to mineral soil compounds (e.g. iron/aluminium oxides and hydroxides or clay minerals), whereas in the aquifer reduction to  $\text{Hg}^0$  could be observed.

Boszke et al. (2008) analysed Hg speciation after a  $\text{Hg}^0$  industrial contamination (spill). The main transformations were volatilization to the atmosphere and oxidation to  $\text{Hg}^{\text{II}}$  followed by binding to humic matter and sulfides (Boszke et al., 2008). Indeed, even after high contamination events, very few  $\text{Hg}^0$  persists in the liquid form:  $\text{Hg}^0$  is either rapidly volatilized or more slowly oxidized to  $\text{Hg}^{\text{II}}$  (Biester et al., 2002).

In a study of Hg speciation from historical gold mine tailing, Slowey et al. (2005b) found that  $\text{Hg}^0$  had been oxidized and transformed to mineral and organic sorption complexes, highly insoluble cinnabar ( $\text{HgS}$ ), Hg oxides and Hg chloride. Moreover, colloidal transport of cinnabar was identified as the dominant leaching mechanism in the mine tailings (Slowey et al., 2005b). Lowry et al. (2004) similarly observed colloidal  $\text{HgS}$  as the dominant transported form of Hg from gold mine waste materials.

Other speciation analyses of gold mine tailings illustrate how Hg speciation can vary from one site to another. In one instance, thermodynamic calculations indicated that >99.8% of mobilized Hg was in the form of Hg-CN complexes (Shaw et al., 2006). Palmieri et al. (2006) found that most of the mercury in a gold mining area was associated with organic matter and sulfur.

In samples from a mercury mining area, Biester et al. (1999) found differences in speciation depending on the time of contamination: older incomplete ore roasting gave more cinnabar than younger tailings in which mineral sorbed  $\text{Hg}^0$  (see section 3.2.1) and unbound  $\text{Hg}^0$  predominated. Studying speciation in another mercury mining area, Navarro et al. (2006) indicated that predominant Hg species in soils and mine wastes was cinnabar ( $\text{HgS}$ ), mainly formed from the weathering of Hg-rich pyrite and elemental  $\text{Hg}^0$  in the matrix, whereas in calcines and tailings the dominant species was  $\text{Hg}^0$ . In the area of the former Almadén (Spain) mercury mining industry, fractionation analyses of soil samples showed predominant Hg sulfides, then associated with Fe-Mn oxyhydroxides and with organic matter, and traces of  $\text{Hg}^0$  (Millan et al., 2011). Land use influence on Hg speciation was also indicated by different speciation results in agricultural and pastoral areas (Millan et al., 2011).

Lechler et al. (1997) studied Hg speciation in soils contaminated by former gold and silver mines. They found that after dissolution of the source  $\text{Hg}^0$ , deeper, high-sulfur soil (reducing) horizons appeared to fix a significant part of the mercury as  $\text{HgS}$ .

Bernaus et al. (2006) assessed Hg speciation in soils polluted by Hg<sup>0</sup> wastes from a chlor-alkali plant. Speciation was observed to quite insoluble inorganic compounds: cinnabar and metacinnabar (HgS), corderoite (Hg<sub>3</sub>S<sub>2</sub>Cl<sub>2</sub>), and small quantities of HgSO<sub>4</sub> and HgO. Similar speciation results were found by Santoro et al. (2010) and Terzano et al. (2010) at a different site contaminated by a chlor-alkali plant. HgO(s) and HgCl<sub>2</sub>(s) are typically species that are sometimes found in highly contaminated soils but not under background conditions (Skylberg, 2010).

Renneberg and Dudas (2001) studied Hg<sup>0</sup> speciation in the presence of co-contamination by hydrocarbons. They found that over time Hg has transformed to HgII and is mostly sorbed to soil organic and mineral materials. Association of Hg with hydrocarbon was significant in one of the five analysed samples (Renneberg and Dudas, 2001).

Bessinger and Marks (2010) investigated soil and cinder contamination in a site of historical mercury fulminate (Hg(CNO)<sub>2</sub>) production. Elemental mercury was not found, and methylmercury was detected at low level. Mercury sulfides were detected in both samples (soil and cinder), and other forms of mercury (i.e. adsorbed Hg) were likely present but not analysed. Bessinger and Marks (2010) further developed a geochemical model for simulating in-situ remediation by incorporation of activated carbon (see report Part 2 – Conceptual and Mathematical model for Mercury Fate and Transport in Soil Systems).

Gray et al. (2006) showed that methylation and demethylation (see section 2.2) processes were significant in mercury mine tailings. Methylmercury concentrations were positively correlated with Hg<sup>2+</sup>, organic carbon and total sulfur concentrations. Different conclusions were drawn in study of soils contaminated by a former chlor-alkali plant. Methylmercury concentrations in soil were negatively correlated to total Hg concentrations, organic carbon and total sulfur concentrations (Remy et al., 2006). High methylmercury/Hg<sub>tot</sub> ratios were mainly related to a combination of low Hg<sub>tot</sub>, low organic C/N ratios and relatively low pH (Remy et al., 2006). Frohne et al. (2011) studied methylation in floodplain soils contaminated by the textile and metal industries. They suggested that the dissolved organic carbon (DOC)/Hg<sub>tot</sub> ratio might be a more important factor for the Hg net methylation (with a positive correlation) than DOC alone. Other important factors influencing methylation were changes in DOC, sulfur cycle, and microbial community structure, whereas Eh and pH values, dissolved Fe<sup>3+</sup>/Fe<sup>2+</sup> and Cl<sup>-</sup> concentrations seemed to play a minor role (Frohne et al., 2011).

## 2 Aqueous speciation of mercury

The main dissolved Hg species in soil systems is  $\text{Hg}^{\text{II}}$  complexes with various inorganic and organic ligands (Loux, 1998; Skyllberg, 2012). Dissolved  $\text{Hg}^0$  and organic Hg forms (mainly monomethylmercury and dimethylmercury) are also present (Ullrich et al., 2001). Methylmercury forms are present in small amounts in soils, but are very important in terms of toxicity because they have the potential to bioaccumulate owing to their lipophilic and proteinbinding properties (Schlüter, 2000; Ullrich et al., 2001). Factors controlling Hg speciation are pH, ionic strength, redox potential, and concentrations of dissolved organic matter (DOM), dissolved oxygen, sulfide and suspended solids in solution. The first three (pH, ionic strength, redox potential) are by far the most important (Gabriel and Williamson, 2004).

Water solubility of Hg compounds varies, e.g.  $\text{CH}_3\text{HgCl}$  and  $\text{CH}_3\text{HgOH}$  more soluble than  $\text{HgCl}_2$  and  $\text{Hg}(\text{OH})_2$  (Gabriel and Williamson, 2004). Highest water solubility occurs in oxygenated environment  $E_h = 350\text{-}400$  mV; this is typical of most soils (Schuster, 1991).

Section 2.1 presents oxido-reduction reactions, while section 2.2 treats the formation and degradation of methylmercury. Sections 2.3 and 2.4 give an overview of (methyl)mercury speciation with inorganic and organic ligands, respectively

### 2.1 Mercury oxidation and reduction

In soil systems, depending on redox conditions, mercury can exist in three valence states:  $\text{Hg}^0$ ,  $\text{Hg}^{\text{I}}$  and  $\text{Hg}^{\text{II}}$  (Gabriel and Williamson, 2004). In the presence of dissolved ligands and organic matter,  $\text{Hg}^{\text{I}}$  and  $\text{Hg}^{\text{II}}$  exist as free species  $\text{Hg}_2^{2+}$  and  $\text{Hg}^{2+}$  but also in many complexed forms. However, most  $\text{Hg}_2^{2+}$  compounds, when exposed to air and light, will oxidize to  $\text{Hg}^{2+}$  (Davis et al., 1997). Therefore, the presence of  $\text{Hg}^{\text{I}}$  in soils is negligible compared to  $\text{Hg}^0$  and  $\text{Hg}^{\text{II}}$ , and the latter is dominant (Schlüter, 2000).

The most widely noted Hg reaction in soils is the reduction of  $\text{Hg}^{2+}$  to  $\text{Hg}^0$  (Gabriel and Williamson, 2004). Reduction to  $\text{Hg}^0$  is usually the first step of mercury volatilization from soil to atmosphere (see section 3.3).  $\text{Hg}^{2+}$  reduction in soils can be biotic or abiotic, and takes place more easily in lower or saturated horizons which are generally reducing environments (Schlüter, 2000).

Abiotic reduction can be mediated by reductants such as dissolved organic matter (humic and fulvic acids) or  $\text{Fe}^{2+}$  (Gabriel and Williamson, 2004; Schlüter, 2000). Mercury reduction by humic and fulvic acids was shown to be affected by solution pH, concentrations of dissolved oxygen, and chloride (Ravichandran, 2004).

Abiotic reduction is influenced by adsorption, because free  $\text{Hg}^{2+}$  in solution has a higher potential for reduction than an adsorbed Hg compound (Mishra et al., 2011; Schlüter, 2000; Schuster, 1991). As a consequence, the reduction of  $\text{Hg}^{2+}$  by reaction with dissolved organic matter (DOM) in the soil upper layers may benefit from the availability of recently deposited atmospheric  $\text{Hg}^{2+}$ , which has not yet been incorporated into the soil matrix and is still relatively available (Schlüter, 2000).

Abiotic reduction of  $\text{Hg}^{2+}$  (and  $\text{CH}_3\text{Hg}^+$ ) to  $\text{Hg}^0$  can also result from photochemical reactions in the first few millimeters of soil (Gabriel and Williamson, 2004; Ravichandran, 2004).

Biotic reduction of  $\text{Hg}^{2+}$  to  $\text{Hg}^0$  can be mediated by various microorganisms (Gabriel and Williamson, 2004). Similarly to abiotic reduction, an increase in DOM inhibits biotic reduction due to the relative unavailability of adsorbed Hg compounds.

## 2.2 Mercury methylation and demethylation

(Mono)methylmercury (MMHg) and dimethylmercury (DMHg) are the most common methylated forms synthesized by microbial activity (Davis et al., 1997). DMHg is practically insoluble in water and is highly volatile, while MMHg has a much higher water solubility (Gabriel and Williamson, 2004).

Generally, the concentration of MMHg is less than 2% of the total mercury concentration in soil (Schlüter, 2000), but this percentage can be higher in wetland or riparian zones (EPA, 1997; Skyllberg et al., 2003). There seems to be an increase in the percentage of MMHg as one goes further from the source of Hg deposition (Skyllberg et al., 2003; Ullrich et al., 2001). The percentage of MMHg compared to the total Hg is generally higher in the aqueous phase compared to the solid phase (Skyllberg, 2012).

Factors controlling methylation and demethylation of Hg have often been studied in aqueous systems such as wetlands, rivers and lakes; because these are the proximate pools of methylmercury before bioaccumulation in the food chain. Ullrich et al. (2001) provided an extensive review of mercury methylation in aqueous systems. Some points of their review are reported here, because the soil solution can be considered as an aqueous system but it should be reminded that most of the literature they cite is specific to surface water.

Mercury methylation can occur in both aerobic and anaerobic environments, but in sediments biotic methylation and MMHg stability appear to be enhanced under anaerobic conditions (Ullrich et al., 2001).

The concentration of bioavailable  $Hg^{II}$  (rather than the total Hg pool) is a primary factor controlling methylation (Ullrich et al., 2001). For example in a study of arctic soils over one summer period, Oiffer and Siciliano (2009) found a net loss of MMHg because most of the mercury seemed to be complexed with DOM and hence unavailable for methylation.  $Hg^{2+}$  dissolved in soil solution is more available for methylation processes than adsorbed  $Hg^{2+}$  (Gabriel and Williamson, 2004; Skyllberg et al., 2003). Using isotope tracers, Jonsson et al. (2012) confirmed that methylation rate can vary a lot depending on the  $Hg^{II}$  species. Methylation rate constants in estuarine sediments were ranked as follows: metacinnabar < cinnabar <  $Hg^{II}$  reacted with mackinawite (Fe,Ni sulfide mineral) <  $Hg^{II}$  bounded to DOM <  $Hg(NO_3)_2(aq)$ .

The efficiency of microbial Hg methylation depends also on microbial activity, which in turn is influenced by temperature, pH and redox potential. Moderately high temperatures likely stimulate microbial activity and contribute to explain higher MMHg production rates usually observed during summer in aqueous systems. Besides, enhanced rates of MMHg production (in aqueous systems) seem to be associated with low pH and the presence of decomposable organic matter in reducing environments (Ullrich et al., 2001). For example, Munthe et al. (2001) found more rapid methylation in wetland soils than in dry forest soils.

The influence of organic matter on Hg methylation is not fully elucidated and may have conflicting effects. On the one hand, organic carbon can enhance methylation by stimulating microorganisms or through direct abiotic methylation of Hg by humic or fulvic substances (Ullrich et al., 2001). On the other hand, high DOM concentrations can decrease the bioavailability of Hg to bacteria particularly in the neutral pH range (Ullrich et al., 2001).

It is important to note that due to demethylation, MMHg concentrations reflect net methylation rather than actual rates of MMHg production (Ullrich et al., 2001). To our knowledge, no studies have examined demethylation in soils. In aqueous systems, MMHg is kinetically stable but is efficiently degraded by microbial action and can also be decomposed by photochemical reactions. MMHg degradation by bacteria mainly involves  $Hg^{II}$  reduction to  $Hg^0$  and appears to be favored under aerobic conditions (Ullrich et al., 2001). However, oxidative demethylation – up to 8-10 % per day – in sediments (and production of  $CO_2$ ) has also been established (Hines et al., 2000; Ullrich et al., 2001).

## 2.3 Inorganic complex forms

The main factors affecting inorganic complex formation are pH, redox conditions, dissolved  $\text{OH}^-$ ,  $\text{Cl}^-$  and  $\text{S}^-$  ions and DOM. Due to their abundance and high stability with  $\text{Hg}^{\text{II}}$ , the main inorganic Hg-complexes are with chloride and hydroxyl (Gabriel and Williamson, 2004).

In oxidized soil conditions the predominant inorganic mercury species are  $\text{Hg}(\text{OH})_2$ ,  $\text{HgCl}_2$ ,  $\text{HgOH}^+$ ,  $\text{HgS}$ , and  $\text{Hg}^0$ .  $\text{Hg}(\text{OH})_2$  and  $\text{HgCl}_2$  are the most frequent (see also Figure 2(a)) due to high abundance and stability of  $\text{OH}^-$  and  $\text{Cl}^-$  (Gabriel and Williamson, 2004).  $\text{HgCl}_2$  is more easily transported in soils than the other inorganic mercury species (less sorption), and typically serves as the substrate for mercury methylation (Han et al., 2006). Mercuric oxide ( $\text{HgO}$ ) can also be encountered in contaminated environments (Ullrich et al., 2001).

In reduced conditions, mercury usually complexes with sulfides to form compounds such as  $\text{HgS}_2^{-2}$ ,  $\text{HgS}(\text{H}_2\text{S})_2$ ,  $\text{Hg}(\text{SH})_2$ ,  $\text{HgSH}^+$ ,  $\text{HgOHSH}$ , and  $\text{HgClSH}$  (Gabriel and Williamson, 2004; Skyllberg, 2012; Ullrich et al., 2001). Relatively insoluble cinnabar ( $\text{HgS}$ ) can also be formed (Skyllberg, 2010) eventually leading to  $\text{HgS}$  precipitation (see also section 3.1.1). The dominant factor controlling the speciation of mercury in the presence of sulfide is the redox potential, which determines whether sulfur is in a reduced or oxidized form (Gabriel and Williamson, 2004). In reducing conditions,  $\text{S}^{2-}$  is normally present in large quantities; therefore  $\text{HgS}$  is mainly formed under reducing conditions (Gabriel and Williamson, 2004; Schuster, 1991). Accumulation of stable  $\text{HgS}$  will result in highly insoluble cinnabar or meta-cinnabar, which further precipitate. However, competing ions ( $\text{Cl}^-$ , DOC,  $\text{Fe}^{2+}$ ) can prevent  $\text{HgS}$  formation (Gabriel and Williamson, 2004; Skyllberg, 2012).

The speciation characteristics of MMHg are similar to those of  $\text{Hg}^{2+}$ .  $\text{CH}_3\text{Hg}^+$  has the following order of affinities for common inorganic ligands:  $\text{SH}^- > \text{OH}^- > \text{Cl}^-$ . In neutral pH range the following forms predominate due to ion abundance:  $\text{CH}_3\text{HgCl} > \text{CH}_3\text{HgOH} > \text{free CH}_3\text{Hg}^+$  (Gabriel and Williamson, 2004).

## 2.4 Organic complex forms

Hg and MMHg complexation with dissolved organic matter has been extensively studied (e.g. Haitzer et al., 2002; 2003; Khwaja et al., 2010; Ravichandran, 2004).

In soils, strong interactions between mercury and organic matter are observed. Mainly the sulfur-containing functional groups (especially thiol groups) are considered to be important for Hg and MMHg binding to DOM (Gabriel and Williamson, 2004; Ravichandran, 2004; Skyllberg, 2012). The thermodynamic calculations of Skyllberg (2012; Table 7.2) including common organic and inorganic ligands found Hg complexes with thiol groups by far the dominant aqueous species in freshwater and wetland soils.

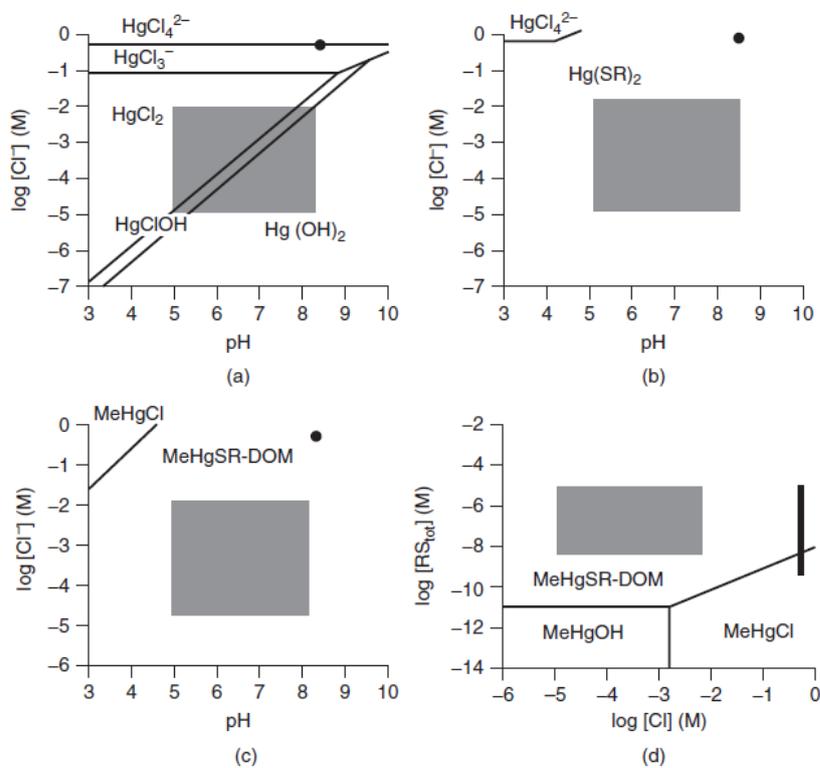
In soils not contaminated by anthropogenic sources strong binding sites in organic matter exceed by far the amount of mercury available (Ravichandran, 2004) even if it assumed that only about 2% of the reduced sulfur is available for binding with mercury (Haitzer et al., 2002). The complexation of Hg to NOM, except under extreme conditions of contamination, outcompetes  $\text{Hg}(\text{OH})_2(\text{aq})$  and  $\text{HgCl}_2(\text{aq})$  (Skyllberg, 2012). Besides, it is often the case that inorganic Hg species, although quite soluble, are further complexed with organic ligands (mainly fulvic and humic acids) and mineral colloids (EPA, 1997; Gabriel and Williamson, 2004).

Even in the presence of sulfide (for which Hg has a strong affinity), some interaction is observed between Hg and DOM, possibly reflecting the formation of a DOM-Hg-sulfide complex or the hydrophobic partitioning of neutral Hg-sulfide complexes into higher-molecular-weight DOM (Miller et al., 2007).

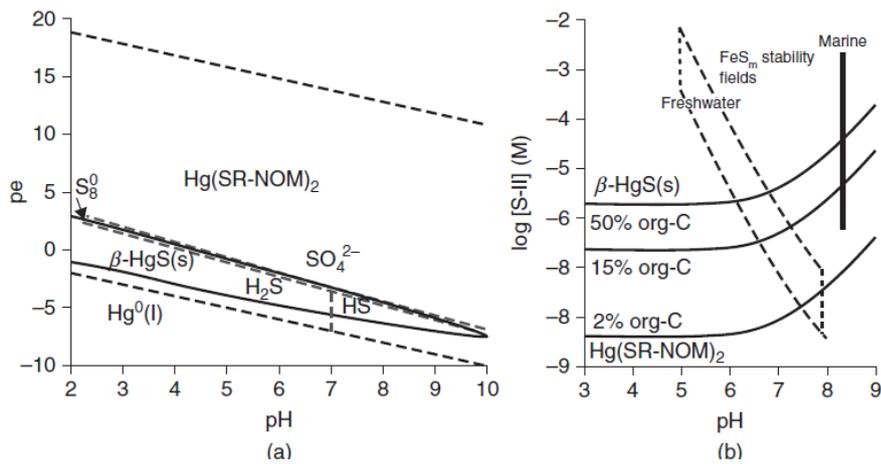
As illustrated in Figure 2(b) for oxic conditions, two-coordinated  $\text{Hg}^{2+}$  complex with organic thiols in DOM is probably the dominant form of mercury organic complexation (Hesterberg et al., 2001; Skyllberg, 2012). Three or four-coordinated complexes may also form under neutral and alkaline pH and high S/Hg ratios (Skyllberg, 2012). Concerning MMHg, the dominant form is the one-coordinated complex MMHg-thiol (see Figure 2(c)).

In case of extreme (industrial) Hg contamination, if Hg is in excess compared to DOM sulfur-containing groups it may then bound to oxygen-containing functional groups such as carboxyl or phenol groups (Reddy and Aiken, 2001; Xia et al., 1999).

Figure 3 shows the dominance diagram of Hg species in soils and sediments as a function of pe and pH (Skyllberg, 2012). This diagram differs from traditionally published dominance diagrams by including Hg complexed by thiol groups associated with DOM, and it can be noticed that the  $\text{Hg}(\text{SR-NOM})_2$  complex dominates over the whole pH scale (Skyllberg, 2012).



**Figure 2 – Dominant Hg aqueous species under oxic conditions (a) in the absence and (b) in the presence of 10 nM of low molecular mass thiols (RS), and (c) for MeHg under oxidized conditions in the presence of DOC 50 mg/L (having 2.3  $\mu\text{M}$  of thiol functional groups, DOM-RS) and (d) a pH of 7. Marine conditions are indicated by black areas, and freshwater conditions by gray areas. Source : Skyllberg (2012).**



**Figure 3 – Dominance diagrams for soils and sediments with 100 ng Hg/g dry mass and 90 mass% pore water content. (a) The solid phase contains 15% organic C (NOM having a concentration of thiol functional groups corresponding to 0.8 mM on a pore water basis): the concentration of inorganic sulfur is 0.1 mM. Solid lines show the stability of mercury species, black broken lines show the stability of water and grey broken lines show the stability of sulfur species. (b) Limiting concentrations of  $S(-II)$  ( $= [H_2S] + [HS^-]$ ) for the precipitation of metacinnabar in sediments with different concentrations of organic C. Regulation of  $S(-II)$  by the solubility of  $FeS_m$  is indicated at freshwater (area bordered by dotted line) and marine conditions (black field). The stability of  $FeS_m$  is in equilibrium with  $Fe^{2+}$  concentrations of  $10^{-3}$  to  $5 \times 10^{-5}$  M in freshwater and  $10^{-6}$  to  $5 \times 10^{-8}$  in marine environments. Source : Skyllberg (2012).**

## 3 Phase transitions of mercury

### 3.1 Dissolution

#### 3.1.1 Dissolution of HgS(s) and HgO(s)

HgS(s) is the common ore of mercury. It is mainly encountered in its most stable crystalline  $\alpha$ -form (cinnabar) or cubic  $\beta$ -form (metacinnabar). Weathering of HgS(s) ore is a common geogenic source of mercury in the environment (section 1.1).

Formation of HgS(s) in soil solution can immobilize mercury, but in the presence of  $\text{Cl}^-$  or DOM, HgS(s) formation is inhibited and solubility and dissolution of HgS(s) are enhanced (Gabriel and Williamson, 2004; Miller et al., 2007; Reddy and Aiken, 2001; Skyllberg, 2012; Waples et al., 2005). Based on the stability constants of Hg-DOM and Hg-sulfide complexes, it is not likely that HgS would be thermodynamically favorable (Miller et al., 2007). Instead, interactions between DOM and HgS surface have been suggested, resulting in the dissolution of HgS and formation of a dissolved Hg-DOM complex (Waples et al., 2005):  $\text{HS-Hg-OH} + \text{HR} = \text{HS-Hg-R} + \text{H}_2\text{O}$ . It has also been suggested that not all DOM types affect HgS(s) dissolution rates in the same way (Gerbig et al., 2012).

Skyllberg (2012) showed that under certain conditions (presence of  $\text{S}_8^0$  and  $\text{pH} > 8$ ) polysulfides increase HgS(s) solubility.

In an experimental study, Waples et al. (2005) found linear dissolution rates of HgS with time. Rates approached constant values at a specific ratio of DOM concentration to cinnabar surface area, suggesting a maximum surface coverage by dissolution-reactive DOM.

Dissolution of montroydite (HgO) was studied by Hocsman et al. (2006) among others. They concluded that montroydite will only occur in nature in highly contaminated sites and that Hg oxidation products formed at the  $\text{Hg}^0(\text{l})/\text{water}$  interface may dissolve rapidly.

#### 3.1.2 Dissolution from non-aqueous liquid form

In case of anthropogenic contamination (see also section 1.2), Hg can be present in the form of a (dense) non-aqueous phase liquid or (D)NAPL, which can further dissolve or volatilize in soil systems.

NAPLs behave differently than dissolved solutes in soil systems. This behavior depends on fluid properties such as interfacial tension, viscosity and density; i.e. properties associated with volatilization and aqueous solubility (Mercer and Cohen, 1990). Mass transfer produces vapor and solute transport in the vadose zone.

Mercury NAPL has a high density and high interfacial tension with water (Devasena and Nambi, 2010). Liquid Hg also has the rare property for a liquid of preferentially wetting air rather than solids (Mercer and Cohen, 1990).

Devasena and Nambi (2010) noted that due to its properties, migration and entrapment of mercury DNAPL cannot be fully described using existing theories and models of DNAPL fate (e.g. Fure et al., 2006; Grant and Gerhard, 2007; Kueper et al., 2003). Models of DNAPL dissolution usually describe the mass transfer via changes in interfacial area as a residual non-wetting phase dissolves. The work of Devasena and Nambi (2010) is to our knowledge the only experimental study devoted to mercury DNAPL in soils, and provides useful inputs for two phase flow models specific to mercury-water systems (see also Annex 6.1). Tsakiroglou and Payatakes (1998) also studied mercury intrusion and capillary pressure curves in porous media, but using artificial glass pore networks.

## 3.2 Sorption

### 3.2.1 Types of sorbent

The literature on mercury adsorption focuses on  $\text{Hg}^{2+}$  because it has a high reactivity and high solubility (Gabriel and Williamson, 2004). However, since the dominant  $\text{Hg}^{\text{II}}$  species in solution are uncharged complexes, the dominant mechanism for sorption on the solid phase is stable insoluble inorganic and organic complex formation rather than ion exchange (Schuster, 1991).

Sorbents can be mineral or organic surfaces. Sorbents can also be categorized as particulate ( $>0.45 \mu\text{m}$ ) and colloidal ( $<0.45 \mu\text{m}$  and  $>1 \text{ nm}$ ) sorbents (Aiken et al., 2011; Gabriel and Williamson, 2004). Due to their large surface area, the finest particles (i.e. colloidal particles) have the highest adsorption capacities (Gabriel and Williamson, 2004). It should be noted that all fulvic acids (i.e. humic acids having a lower molecular weight and higher oxygen content) are colloids according to the International Humic Substances Society.

Sorption capability of  $\text{Hg}^{2+}$  is highest for organic matter in the particulate and dissolved phases (especially humic and fulvic substances). In conditions of low Hg/DOM ratio (i.e. no high Hg contamination), sorption of Hg to DOM is controlled by a small fraction of DOM molecules containing a reactive thiol functional group (Haitzer et al., 2002; Ravichandran, 2004).

Other potentially important sorbents are oxides, hydroxides, oxyhydroxides of Fe/Mn/Al (best sorbents  $\text{MnOOH}$  and  $\text{FeOOH}$ ), amorphous  $\text{FeS}$  under reducing conditions, and clay minerals (Gabriel and Williamson, 2004). However, except under highly contaminated conditions and/or low organic matter environments, Hg and MMHg will not directly bind to oxygen functional groups (Skylberg, 2012). Therefore, when using Hg/DOM distribution coefficients for modeling the fate and transport of Hg, it is important to take into account the Hg/DOM ratios at which they were determined. For example, at Hg/DOM ratios  $<1 \mu\text{g}$  of Hg/mg of DOM, Haitzer et al. (2002) observed very strong Hg-thiol bonds. At Hg/DOM ratios  $>10 \mu\text{g}$  of Hg/mg of DOM (as used in most laboratory studies), much lower distribution coefficients are found, indicative of Hg binding mainly to oxygen (carboxyl and phenol) functional groups (Drexel et al., 2002; Haitzer et al., 2002).

Detailed modeling of  $\text{Hg}^{2+}$  binding to DOM showed the existence of bidentate binding by one thiol group and one other group (statistical mixture of functional groups containing oxygen, nitrogen or sulfur; Haitzer et al., 2003), and indirect adsorption can occur through  $\text{Hg}(\text{SR})_2$  or  $\text{MMHg}(\text{SR})$  forming ternary complexes with reactive  $-\text{XOH}$  groups (Senevirathna et al., 2011; Skylberg, 2012).

Although highly volatile (see section 3.3), elemental mercury ( $\text{Hg}^0$ ) can also be adsorbed to soil material and has a stronger sorption affinity for DOM than for minerals (metal oxides) (Schlüter, 2000).  $\text{Hg}^0$  adsorption increases with sorbent surface area, but the physical structure of soils can also play a role, for example through restricted migration in clay horizons increasing the contact time between  $\text{Hg}^0$  and the soil matrix and therefore increasing adsorption (Schlüter, 2000).

In a study of Hg distribution in soil profiles near chlor-alkali plants, migration of Hg to deeper soil layers was suggested to be most effective if Hg was bound to soluble organic complexes (cf. colloid-facilitated transport), whereas Hg bound to mineral components or weak Hg complexes (e.g.  $\text{Hg}^0$  bound to inner-sphere sorption sites) were effectively retained in the uppermost soil layer (5 cm) (Biester et al., 2002).

Sorption to colloids is very important to understand the fate of mercury in soils, because colloid-facilitated transport is sometimes referred to as the main Hg transport process (Gao et al., 2011; Slowey et al., 2005a; Slowey et al., 2005b). Interaction between heavy metals and colloids can potentially be of three types : adsorption, precipitation and cation exchange (Gao et al., 2011). However, among these adsorption is the predominant and most influential for heavy metal mobility in soils.

The three main active colloidal constituents in soils are clay minerals, metal oxides and organic matter (humic and fulvic acids). Colloids may also be classified by surface functional groups: surface hydroxyl, carbonyl, or organic complexation group (Gao et al., 2011). Besides, the sorptive behavior of colloidal particles is not the simple sum of the contributions of the single constituents (it is less because of blockage or loss of accessibility of many functional groups; Cruz-Guzmán et al., 2003).

Studying  $\text{Hg}^{2+}$  (at high concentration) sorption on soil humic acid, ferrihydrite and montmorillonite colloidal particles, Cruz-Guzmán et al. (2003) found that sorption was much higher on humic acids than on ferrihydrite and montmorillonite. This can be interpreted as the result of strong sorbed complexes with organic S-containing functional groups and a much higher CEC for humic substances than for these minerals (Gao et al., 2011).

Studying the distribution of colloidal Hg in soil, Santoro et al. (2012) found a positive correlation between the concentration of Hg in the colloidal fraction and OM content in samples polluted by a former chlor-alkali plant. Rather than attributing this correlation to sorption of Hg on DOM, they suggested that HgS sorption to DOM could prevent the growth of HgS colloids into larger crystals. This would therefore augment Hg colloidal fraction and increase the risk of Hg mobilization. The effect of colloid sorption to DOM was experimentally investigated by Morales et al. (2011) using polystyrene and carboxylated spheres as colloids. They observed that humic acids could electrosterically stabilize colloids and thus enhanced colloid transport significantly, while fulvic acids hardly affected transport.

More information on colloid-facilitated transport in soils in Flury and Qiu (2008) and in Gao et al. (2011).

### **3.2.2 Effect of dissolved species ( $\text{S}^{2-}$ , $\text{Cl}^-$ and DOC), pH and heat**

Chloride has been shown to interfere with  $\text{Hg}^{2+}$  adsorption in soils. In general (but not always), an increase in  $\text{Cl}^-$  concentration reduces adsorption of  $\text{Hg}^{2+}$  and  $\text{CH}_3\text{Hg}^+$  to inorganic and organic adsorbents (Gabriel and Williamson, 2004). Similarly, mercury forms a covalent bond with  $\text{OH}^-$ . This highly stable dissolved complex minimizes the bond between the mineral surface (oxide) and  $\text{OH}^-$ . As a result, the overall mercury adsorption to mineral surfaces decreases (Schuster, 1991).

The formation of mercury-DOC complexes has an impact on Hg adsorption as generally known for heavy metals, but contrasting effects have been found for mercury (Schuster, 1991). On the one hand, a decrease in Hg adsorption to mineral surfaces could be due to the strong DOC-Hg complex stability that prevents mercury from adsorbing to the mineral surface. On the other hand, an increase in Hg adsorption could be due to a high affinity of the dissolved organic ligand for the mineral surface (Gabriel and Williamson, 2004; Schuster, 1991).

Mercury adsorption is also influenced by soil pH (Gabriel and Williamson, 2004). When pH is <4.5-5, organic matter is the most efficient mercury sorbent. In neutral to alkaline soil, mineral components are more efficient sorbents. As with all metals, adsorption generally decreases with decreasing pH, due to  $\text{H}^+$  removing and replacing Hg ions (Gabriel and Williamson, 2004). However, it has sometimes been stated that mercury adsorption to humic matter increases at lower pH (e.g. Jing et al., 2007; Sarkar et al., 2000).

Heat has been shown to decrease  $\text{Hg}^{2+}$  adsorption (Biester and Scholz, 1996). However, in terms of natural occurring temperatures, heat influence on mercury desorption was found only for  $\text{HgCl}_2$  (Biester and Scholz, 1996; Gabriel and Williamson, 2004).

### **3.2.3 Equilibrium or kinetic sorption, hysteresis**

Kinetic studies, such as that reported by Liao et al. (2009), showed that between 24 and 48h, Hg adsorption reached a quasi-equilibrium state. Amacher et al. (1990) stated that generally multiple

reaction kinetics (e.g. rapid and slow reversible reactions and irreversible reaction) were needed to fit  $\text{Hg}^{2+}$  sorption.

In a study on four loamy soils at pH 6, Yin et al. (1997b) found that both  $\text{Hg}^{2+}$  adsorption and desorption were characterized by a biphasic pattern, a fast step (a few minutes) followed by a slow step (<24 h in total). Besides, adsorption and desorption rate coefficients were inversely correlated with the soil organic C content.

In their study on three different soils, Liao et al. (2009) found rapid and nonlinear  $\text{Hg}^{2+}$  adsorption with the sorption capacities for clay > loam > sand. A multi-reaction model (based on Zhang and Selim, 2006) including sequential and parallel equilibrium and/or sorption reactions was also used to fit the adsorption data. Yin et al. (1997a) similarly advocated the use of a multireaction model to describe Hg retention in soils, justified by the many different solid phases that can immobilize Hg. Indeed, even in the absence of soil organic matter, Hg transport can be highly retarded due to adsorption to mineral sorbents (Jing et al., 2008). This was confirmed by Liao et al. (2009) who similarly found significant Hg adsorption in the absence of soil organic matter.

S-type isotherms have also been proposed to fit experimental data of Hg adsorption, characterized by limited sorption at low initial concentrations followed increasing adsorption as Hg concentration in solution increases (Yin et al., 1997a). Isotherms of the S-type may be due to the early complexation of Hg by DOM. When  $\text{Hg}^{2+}$  concentration exceeds DOM complexation capacity, adsorption to soil particle surface becomes competitive (Yin et al., 1997a).

In a review of adsorption isotherms of heavy metals and other chemicals, Ho and McKay (1999) found that pseudo-first order model fitted well the initial period of the adsorption reaction, but that pseudo-second order model provided the best fit on a longer period. Recently, the question on how these pseudo-first and second order models can be interpreted in view of the Langmuir theoretical model has been debated (Liu and Shen, 2008; Marczewski, 2010; Plazinski et al., 2009; Saeid, 2004).

Hysteresis of mercury adsorption/desorption in soils has frequently been demonstrated (e.g. Jing et al., 2008; Liao et al., 2009; Miretzky et al., 2005; Yin et al., 1997b). Besides, the fraction of Hg irreversibly sorbed was shown to be correlated with soil organic content (e.g. Jing et al., 2007; Liao et al., 2009). It has been suggested that irreversible sorption may be due to diffusion of  $\text{Hg}^{\text{II}}$  through intraparticle micropores of soil organic matter (Jing et al., 2007; Yin et al., 1997b), and to high affinity for S-containing groups (Yin et al., 1997b).

In the study of Jing et al. (2007), the influence of pH on  $\text{Hg}^{2+}$  desorption displayed a “U” pattern; i.e. a decrease with increasing pH in the lower pH range (3-5) and an increase with increasing pH for  $\text{pH} > 7$ . Strong adsorption of metal hydroxo complexes ( $\text{HgOH}^+$  and  $\text{Hg}(\text{OH})_2^0$ ), hydrolysis of Al on exchange sites, competition of protons for adsorbing sites, and acid catalyzed dissolution of reactive oxide sites were suggested to explain this “U” pattern (Jing et al., 2007).

$\text{CH}_3\text{Hg}^+$  is less strongly adsorbed to soil than  $\text{Hg}^{2+}$  (Gabriel and Williamson, 2004).

### 3.3 Volatilization

Volatilization of mercury from soils has been extensively studied, but it still represents the major uncertainty in global/regional annual budget of non point source mercury transport (Grigal, 2002).

Volatilization is assumed to require three steps: (i) reduction of  $\text{Hg}^{\text{II}}$  to  $\text{Hg}^0$ , (ii) diffusion or mass transport of  $\text{Hg}^0$  to the soil surface (in gaseous and aqueous phases), and (iii) diffusion or mass transport of the  $\text{Hg}^0$  across the soil–air boundary layer into the atmosphere (Schlüter, 2000). DMHg is also volatile and can contribute to mercury volatilization from soils, while MMHg and  $\text{Hg}^{\text{II}}$  salts are of minor importance.  $\text{CH}_3\text{HgOH}$  and  $\text{CH}_3\text{HgCl}$  are volatile compounds, but MMHg volatility decreases due to the high affinity to solute and solid organic matter (Schlüter, 2000).

Sunlight (UV radiation) and heat emitted from sunlight (air and soil temperature) are considered the main factors stimulating  $\text{Hg}^0$  volatilization (through enhanced reduction of  $\text{Hg}^{2+}$  to  $\text{Hg}^0$ ) (Gabriel and Williamson, 2004). Looking at the kinetics of Hg emission data, Quinones and Carpi (2011) suggested that mercury emissions from soil samples may follow a multicompartment model (driven by exposure to light) in which one compartment of the emissions spectrum is limited to the surface (<2 cm) of the soil matrix and a second is driven by a subsurface phenomenon.

Apart from sunlight and heat, all factors having an impact on  $\text{Hg}^{\text{II}}$  reduction (section 2.1) are also expected to influence volatilization. For instance,  $\text{Hg}^{2+}$  volatilization decreases with increasing DOM/ $\text{Cl}^-$  content, and decreasing moisture content (Schlüter, 2000). Soil moisture content is generally found to be very important to determine the volatilization rate (Schlüter, 2000). Generally, mercury volatilization increases with increasing water content up to a certain level (Rinklebe et al., 2010; Song and Van Heyst, 2005).

Fritsche et al. (2008) suggested that biotic processes are partially controlling  $\text{Hg}^0$  volatilization in uncontaminated soils. Biotic action could be direct (biotic reduction of  $\text{Hg}^{2+}$ ) or indirect (microbiological degradation of organic matter followed by  $\text{Hg}^{2+}$  abiotic reduction). Biological reduction of  $\text{Hg}^{2+}$  to  $\text{Hg}^0$  is favoured in soils of high  $\text{Hg}^{2+}$  availability and microbiological activity. Abiological reduction is favoured in soils of high organic matter content and/or low Eh. Microbiological reactions can cause considerably higher mercury evaporation rates than abiotically mediated evaporation (Schlüter, 2000).

In a study of uncontaminated (background) forest soils, soil temperature, redox conditions and organic matter were significant factors affecting gaseous mercury concentrations (Moore and Castro, 2012).

Other factors influencing volatilization – via point (ii) above – are soil physical characteristics, sorption capacity, mercury species and content and pH (Schlüter, 2000).

Finally, meteorological conditions (wind speed, relative humidity, turbulent mixing of air layers, etc.) can affect Hg volatilization by limiting or stimulating transport and release – points (ii) and (iii) above (Gabriel and Williamson, 2004; Schlüter, 2000). Meteorological factors may become especially important in Hg enriched or contaminated areas, while in background areas formation of  $\text{Hg}^0$  (or DMHg) will be the rate limiting factor (Schlüter, 2000).

Values of mercury emission rates from background areas reported in the literature are usually  $<0.2 \mu\text{g m}^{-2} \text{h}^{-1}$  and significantly smaller than from mercury enriched or contaminated soils (Schlüter, 2000; Schroeder and Munthe, 1998; Schroeder et al., 1989). Emissions from uncontaminated urban soils varied significantly between different land uses and ranged from  $8.7 \times 10^{-4}$  to  $4.5 \times 10^{-3} \mu\text{g m}^{-2} \text{h}^{-1}$  (Gabriel et al., 2005). For comparison, Rinklebe et al. (2010) measured 0.01 to  $0.85 \mu\text{g m}^{-2} \text{h}^{-1}$  Hg emission from heavily polluted floodplain soils. Volatilization losses in the subsurface from NAPLs can also be expected, especially where NAPL is close to the surface or in dry sandy soils (Mercer and Cohen, 1990).

Transport of gaseous mercury can also be significant laterally. Walvoord et al. (2008) observed long-distance (several tens or hundreds of meters) migration of elemental mercury released from a buried landfill and transported through the unsaturated zone.

## 4 Conclusion

Sources of mercury in soil systems include atmospheric deposition, geogenic sources and anthropogenic contamination. Sites of anthropogenic contamination are characterised by high Hg contamination and the presence of NAPL is even possible. The present literature review indicates that in the case of highly contaminated soils many processes occurring in background soils are altered or influenced by local factors (e.g. the presence of specific species, high Hg concentrations, the presence of a NAPL or co-contaminants).

Mercury sinks include volatilization to the atmosphere, and transport to groundwater or surface water. Mercury is usually characterized by a high affinity for soil mineral and organic particles and long residence time in soils. However, under given conditions transport of dissolved Hg species or colloidal-facilitated transport can pose significant threats of environmental contamination.

Mercury in soils exists in different phases: dissolved or complexed Hg in the aqueous phase, adsorbed to mineral or organic sorbents, in the gas phase or in the form of non-aqueous phase liquid. The present document reviewed factors and processes determining mercury speciation and transformation.

Under oxidized surface soil conditions, Hg and MMHg form almost exclusively complexes with thiols. Common inorganic mercury forms are  $\text{Hg}(\text{OH})_2$ ,  $\text{HgCl}_2$ ,  $\text{HgOH}^+$ ,  $\text{HgS}$  and  $\text{Hg}^0$ . In reduced environments common mercury forms are  $\text{HgSH}^+$ ,  $\text{HgOHSH}$ , and  $\text{HgClSH}$ . These mercury forms are generally bound to organic and mineral ions/molecules.

Sorption capability of  $\text{Hg}^{2+}$  is highest for organic matter in the particulate and dissolved phases (especially humic and fulvic substances). Adsorption rate depends mainly on initial  $\text{Hg}^{2+}$  concentration, organic matter content, ions in solution. Hysteresis of mercury adsorption/desorption in soils is generally recognized. Sorption to colloids is very important because colloidal-facilitated transport has been referred to as the main Hg transport process.

Volatilization is a three-step process.  $\text{Hg}^{\text{II}}$  is first reduced to  $\text{Hg}^0$ . Then,  $\text{Hg}^0$  is transported/diffused to the soil surface, and finally volatilized across the soil–air boundary layer into the atmosphere. Sunlight and heat are the main factors driving Hg volatilization.

## 5 References

- Aiken, G. R., Gilmour, C. C., Krabbenhoft, D. P. and Orem, W. (2011), Dissolved Organic Matter in the Florida Everglades: Implications for Ecosystem Restoration, *Critical Reviews in Environmental Science and Technology* 41(sup1): 217-248.
- Amacher, M. C., Selim, H. M. and Iskandar, I. K. (1990), Kinetics of Mercuric Chloride Retention by Soils, *J. Environ. Qual.* 19(3): 382-388.
- Bernaus, A., Gaona, X., van Ree, D. and Valiente, M. (2006), Determination of mercury in polluted soils surrounding a chlor-alkali plant: Direct speciation by X-ray absorption spectroscopy techniques and preliminary geochemical characterisation of the area, *Analytica Chimica Acta* 565(1): 73-80.
- Bessinger, B. A. and Marks, C. D. (2010), Treatment of mercury-contaminated soils with activated carbon: A laboratory, field, and modeling study, *Remediation Journal* 21(1): 115-135.
- Biester, H., Gosar, M. and Müller, G. (1999), Mercury speciation in tailings of the Idrija mercury mine, *Journal of Geochemical Exploration* 65(3): 195-204.
- Biester, H., Müller, G. and Schöler, H. F. (2002), Binding and mobility of mercury in soils contaminated by emissions from chlor-alkali plants, *Science of The Total Environment* 284(1-3): 191-203.
- Biester, H. and Scholz, C. (1996), Determination of Mercury Binding Forms in Contaminated Soils: Mercury Pyrolysis versus Sequential Extractions, *Environmental Science & Technology* 31(1): 233-239.
- Bloom, N. S., Preus, E., Katon, J. and Hiltner, M. (2003), Selective extractions to assess the biogeochemically relevant fractionation of inorganic mercury in sediments and soils, *Analytica Chimica Acta* 479(2): 233-248.
- Bollen, A., Wenke, A. and Biester, H. (2008), Mercury speciation analyses in HgCl<sub>2</sub>-contaminated soils and groundwater-Implications for risk assessment and remediation strategies, *Water Research* 42(1-2): 91-100.
- Boszke, L., Kowalski, A., Astel, A., Barański, A., Gworek, B. and Siepak, J. (2008), Mercury mobility and bioavailability in soil from contaminated area, *Environmental Geology* 55(5): 1075-1087.
- Clever, H. (1985), The Solubility of Mercury and Some Sparingly Soluble Mercury Salts in Water and Aqueous Electrolyte Solutions, *Journal of physical and chemical reference data* 14(3): 631.
- Cruz-Guzmán, M., Celis, R., Hermosín, M. C., Leone, P., Nègre, M. and Cornejo, J. (2003), Sorption-Desorption of Lead (II) and Mercury (II) by Model Associations of Soil Colloids, *Soil Science Society of America Journal* 67(5): 1378-1387.
- Davis, A., Bloom, N. S. and Que Hee, S. S. (1997), The Environmental Geochemistry and Bioaccessibility of Mercury in Soils and Sediments: A Review, *Risk Analysis* 17(5): 557-569.
- Devasena, M. and Nambi, I. M. (2010), Migration and entrapment of mercury in porous media, *Journal of Contaminant Hydrology* 117(1-4): 60-70.
- Ding, Z., Wu, H., Feng, X., Liu, J., Liu, Y., Yuan, Y., Zhang, L., Lin, G. and Jiayong, P. (2011), Distribution of Hg in mangrove trees and its implication for Hg enrichment in the mangrove ecosystem, *Applied Geochemistry* 26(2): 205-212.
- Drexel, R. T., Haitzer, M., Ryan, J. N., Aiken, G. R. and Nagy, K. L. (2002), Mercury(II) Sorption to Two Florida Everglades Peats: Evidence for Strong and Weak Binding and Competition by Dissolved Organic Matter Released from the Peat, *Environmental Science & Technology* 36(19): 4058-4064.

- Ebadian, M. A., Marshall, A. and Yong, C. (2001), *Mercury Contaminated Material Decontamination Methods: Investigation and Assessment*, Hemispheric Centre for Environmental Technology, Florida International University, Miami. HCET-2000-DO53-002-04, 73 p.
- EPA (1997), *Mercury Study Report to Congress. Volume III: Fate and Transport of Mercury in the Environment*, US EPA, 376 p.
- Flury, M. and Qiu, H. (2008), Modeling Colloid-Facilitated Contaminant Transport in the Vadose Zone, *Vadose Zone Journal* 7(2): 682-697.
- Fritsche, J., Obrist, D. and Alewell, C. (2008), Evidence of microbial control of Hg<sup>0</sup> emissions from uncontaminated terrestrial soils, *Journal of Plant Nutrition and Soil Science* 171: 200-209.
- Frohne, T., Rinklebe, J., Langer, U., Du Laing, G., Mothes, S. and Wennrich, R. (2011), Biogeochemical factors affecting mercury methylation rate in two contaminated floodplain soils, *Biogeosciences Discussions* 8: 8925-8959.
- Fure, A. D., Jawitz, J. W. and Annable, M. D. (2006), DNAPL source depletion: Linking architecture and flux response, *Journal of Contaminant Hydrology* 85(3-4): 118-140.
- Gabriel, M. and Williamson, D. (2004), Principal Biogeochemical Factors Affecting the Speciation And Transport of Mercury through the terrestrial environment, *Environmental Geochemistry and Health* 26(3): 421-434.
- Gabriel, M., Williamson, D., Brooks, S., Zhang, H. and Lindberg, S. (2005), Spatial variability of mercury emissions from soils in a southeastern US urban environment, *Environmental Geology* 48(7): 955-964.
- Gao, B., Xinde, C., Yan, D., Yongming, L. and Lena, M. Q. (2011), Colloid Deposition and Release in Soils and Their Association With Heavy Metals, *Critical Reviews in Environmental Science and Technology* 41: 336–372.
- Gerbig, C. A., Ryan, J. N. and Aiken, G. R. (2012), *The Effects of Dissolved Organic Matter on Mercury Biogeochemistry*. Environmental Chemistry and Toxicology of Mercury, John Wiley & Sons, Inc.: 259-292.
- Gerhard, J. I., Pang, T. and Kueper, B. H. (2007), Time Scales of DNAPL Migration in Sandy Aquifers Examined via Numerical Simulation, *Ground Water* 45(2): 147-157.
- Grant, G. P. and Gerhard, J. I. (2007), Simulating the dissolution of a complex dense nonaqueous phase liquid source zone: 1. Model to predict interfacial area, *Water Resour. Res.* 43(12): W12410.
- Gray, J. E., Hines, M. E. and Biester, H. (2006), Mercury methylation influenced by areas of past mercury mining in the Terlingua district, Southwest Texas, USA, *Applied Geochemistry* 21(11): 1940-1954.
- Grigal, D. F. (2002), Inputs and outputs of mercury from terrestrial watersheds: a review, *Environmental Reviews* 10(1): 1-39.
- Grigal, D. F. (2003), Mercury Sequestration in Forests and Peatlands: A Review, *Journal of Environmental Quality* 32: 393-405.
- Guedron, S., Grimaldi, C., Chauvel, C., Spadini, L. and Grimaldi, M. (2006), Weathering versus atmospheric contributions to mercury concentrations in French Guiana soils, *Applied Geochemistry* 21(11): 2010-2022.
- Gustin, M. S., Engle, M., Ericksen, J., Lyman, S., Stamenkovic, J. and Xin, M. (2006), Mercury exchange between the atmosphere and low mercury containing substrates, *Applied Geochemistry* 21(11): 1913-1923.
- Haitzer, M., Aiken, G. R. and Ryan, J. N. (2002), Binding of Mercury(II) to Dissolved Organic Matter: The Role of the Mercury-to-DOM Concentration Ratio, *Environmental Science & Technology* 36(16): 3564-3570.

- Haitzer, M., Aiken, G. R. and Ryan, J. N. (2003), Binding of Mercury(II) to Aquatic Humic Substances: Influence of pH and Source of Humic Substances, *Environmental Science & Technology* 37(11): 2436-2441.
- Han, F. X., Su, Y., Monts, D. L., Waggoner, C. A. and Plodinec, M. J. (2006), Binding, distribution, and plant uptake of mercury in a soil from Oak Ridge, Tennessee, USA, *Science of The Total Environment* 368(2-3): 753-768.
- Hesterberg, D., Chou, J. W., Hutchison, K. J. and Sayers, D. E. (2001), Bonding of Hg(II) to Reduced Organic Sulfur in Humic Acid As Affected by S/Hg Ratio, *Environmental Science & Technology* 35(13): 2741-2745.
- Hines, M. E., Horvat, M., Faganeli, J., Bonzongo, J.-C. J., Barkay, T., Major, E. B., Scott, K. J., Bailey, E. A., Warwick, J. J. and Lyons, W. B. (2000), Mercury Biogeochemistry in the Idrija River, Slovenia, from above the Mine into the Gulf of Trieste, *Environmental Research* 83(2): 129-139.
- Ho, Y. S. and McKay, G. (1999), Pseudo-second order model for sorption processes, *Process Biochemistry* 34(5): 451-465.
- Hocsman, A., Di Nezio, S., Charlet, L. and Avena, M. (2006), On the mechanisms of dissolution of montroydite [HgO(s)]: Dependence of the dissolution rate on pH, temperature, and stirring rate, *Journal of Colloid and Interface Science* 297(2): 696-704.
- Jing, Y. D., He, Z. L. and Yang, X. E. (2007), Effects of pH, organic acids, and competitive cations on mercury desorption in soils, *Chemosphere* 69(10): 1662-1669.
- Jing, Y. D., He, Z. L. and Yang, X. E. (2008), Adsorption/Desorption Characteristics of Mercury in Paddy Soils of China, *Journal of Environmental Quality* 37(2): 680-688.
- Jonsson, S., Skyllberg, U., Nilsson, M. B., Westlund, P.-O., Shchukarev, A., Lundberg, E. and Björn, E. (2012), Mercury Methylation Rates for Geochemically Relevant HgII Species in Sediments, *Environmental Science & Technology* 46(21): 11653-11659.
- Khwaja, A. R., Bloom, P. R. and Brezonik, P. L. (2010), Binding Strength of Methylmercury to Aquatic NOM, *Environmental Science & Technology* 44(16): 6151-6156.
- Kueper, B. H., Wealthall, G. P., Smith, J. W. N., Leharne, S. A. and Lerner, D. N. (2003), An illustrated handbook of DNAPL transport and fate in the subsurface. Environment Agency, Bristol, UK.
- Lechler, P. J., Miller, J. R., Hsu, L.-C. and Desilets, M. O. (1997), Mercury mobility at the Carson River Superfund Site, west-central Nevada, USA: Interpretation of mercury speciation data in mill tailings, soils, and sediments, *Journal of Geochemical Exploration* 58(2-3): 259-267.
- Lenhard, R. J., Oostrom, M. and Dane, J. H. (2004), A constitutive model for air-NAPL-water flow in the vadose zone accounting for immobile, non-occluded (residual) NAPL in strongly water-wet porous media, *Journal of Contaminant Hydrology* 71(1-4): 261-282.
- Liao, L., Selim, H. M. and DeLaune, R. D. (2009), Mercury Adsorption-Desorption and Transport in Soils, *Journal of Environmental Quality* 38(4): 1608-1616.
- Lin, C.-J. and Pehkonen, S. O. (1999), The chemistry of atmospheric mercury: a review, *Atmospheric Environment* 33(13): 2067-2079.
- Liu, Y. and Shen, L. (2008), From Langmuir Kinetics to First- and Second-Order Rate Equations for Adsorption, *Langmuir* 24(20): 11625-11630.
- Loux, N. T. (1998), An assessment of mercury-species-dependent binding with natural organic carbon, *Chemical Speciation and Bioavailability* 10(4): 127-136.
- Lowry, G. V., Shaw, S., Kim, C. S., Rytuba, J. J. and Brown, G. E. J. (2004), Macroscopic and Microscopic Observations of Particle-Facilitated Mercury Transport from New Idria and Sulphur Bank Mercury Mine Tailings, *Environmental Science & Technology* 38(19): 5101-5111.

- Marczewski, A. W. (2010), Analysis of Kinetic Langmuir Model. Part I: Integrated Kinetic Langmuir Equation (IKL): A New Complete Analytical Solution of the Langmuir Rate Equation, *Langmuir* 26(19): 15229-15238.
- Marsman, A., Hartog, N., Keijzer, T. and Sweijen, T. (2012), Modelling the DNAPL spreading behavior of pure phase elemental mercury DNAPL in soil and groundwater systems. NICOLE Technical Meeting, Brussels, 3-4 December 2012, NICOLE.
- Mason, R. P., Fitzgerald, W. F. and Morel, F. M. M. (1994), The biogeochemical cycling of elemental mercury: Anthropogenic influences, *Geochimica et Cosmochimica Acta* 58(15): 3191-3198.
- Mercer, J. W. and Cohen, R. M. (1990), A review of immiscible fluids in the subsurface: Properties, models, characterization and remediation, *Journal of Contaminant Hydrology* 6(2): 107-163.
- Millan, R., Schmid, T., Sierra, M. J., Carrasco-Gil, S., Villadóniga, M., Rico, C., Ledesma, D. M. S. and Puente, F. J. D. (2011), Spatial variation of biological and pedological properties in an area affected by a metallurgical mercury plant: Almadenejos (Spain), *Applied Geochemistry* 26(2): 174-181.
- Miller, C. L., Mason, R. P., Gilmour, C. C. and Heyes, A. (2007), Influence of dissolved organic matter on the complexation of mercury under sulfidic conditions, *Environmental Toxicology and Chemistry* 26(4): 624-633.
- Miller, C. T. (1998), Multiphase flow and transport modeling in heterogeneous porous media: challenges and approaches, *Advances in Water Resources* 21(2): 77.
- Miretzky, P., Bisinoti, M. r. C. and Jardim, W. F. (2005), Sorption of mercury (II) in Amazon soils from column studies, *Chemosphere* 60(11): 1583-1589.
- Mishra, B., O'Loughlin, E. J., Boyanov, M. I. and Kemner, K. M. (2011), Binding of HgII to High-Affinity Sites on Bacteria Inhibits Reduction to Hg0 by Mixed FeII/III Phases, *Environmental Science & Technology* 45(22): 9597-9603.
- Moore, C. W. and Castro, M. S. (2012), Investigation of factors affecting gaseous mercury concentrations in soils, *Science of The Total Environment* in press.
- Morales, V. L., Zhang, W., Gao, B., Lion, L. W., Bisogni Jr, J. J., McDonough, B. A. and Steenhuis, T. S. (2011), Impact of dissolved organic matter on colloid transport in the vadose zone: Deterministic approximation of transport deposition coefficients from polymeric coating characteristics, *Water Research* 45(4): 1691-1701.
- Munthe, J., Lyvén, B., Parkman, H., Lee, Y.-H., Iverfeldt, Å., Haraldsson, C., Verta, M. and Porvari, P. (2001), Mobility and Methylation of Mercury in Forest Soils Development of an In-Situ Stable Isotope Tracer Technique and Initial Results, *Water, Air, & Soil Pollution: Focus* 1(3): 385-393.
- Navarro, A., Biester, H., Mendoza, J. L. and Cardellach, E. (2006), Mercury speciation and mobilization in contaminated soils of the Valle del Azogue Hg mine (SE, Spain), *Environmental Geology* 49(8): 1089-1101.
- Obrist, D., Johnson, D. W., Lindberg, S. E., Luo, Y., Hararuk, O., Bracho, R., Battles, J. J., Dail, D. B., Edmonds, R. L., Monson, R. K., Ollinger, S. V., Pallardy, S. G., Pregitzer, K. S. and Todd, D. E. (2011), Mercury Distribution Across 14 U.S. Forests. Part I: Spatial Patterns of Concentrations in Biomass, Litter, and Soils, *Environmental Science & Technology* 45(9): 3974-3981.
- Oiffer, L. and Siciliano, S. D. (2009), Methyl mercury production and loss in Arctic soil, *Science of The Total Environment* 407(5): 1691-1700.
- Oostrom, M., White, M. D., Lenhard, R. J., Van Geel, P. J. and Wietsma, T. W. (2005), A Comparison of Models Describing Residual NAPL Formation in the Vadose Zone, *Vadose Zone J.* 4(1): 163-174.

- Palmieri, H. E. L., Nalini Jr, H. A., Leonel, L. V., Windmüller, C. C., Santos, R. C. and de Brito, W. (2006), Quantification and speciation of mercury in soils from the Tripuí Ecological Station, Minas Gerais, Brazil, *Science of The Total Environment* 368(1): 69-78.
- Pant, P., Allen, M. and Tansel, B. (2010), Mercury Uptake and Translocation in *Impatiens walleriana* Plants Grown in the Contaminated Soil from Oak Ridge, *International Journal of Phytoremediation* 13(2): 168-176.
- Pérez-Sanz, A., Millán, R., Sierra, M. J., Alarcín, R., García, P., Gil-Díaz, M., Vazquez, S. and Lobo, M. C. (2012), Mercury uptake by *Silene vulgaris* grown on contaminated spiked soils, *Journal of Environmental Management* 95, Supplement: S233-S237.
- Plazinski, W., Rudzinski, W. and Plazinska, A. (2009), Theoretical models of sorption kinetics including a surface reaction mechanism: A review, *Advances in Colloid and Interface Science* 152(1-2): 2-13.
- Quinones, J. L. and Carpi, A. (2011), An Investigation of the Kinetic Processes Influencing Mercury Emissions from Sand and Soil Samples of Varying Thickness, *Journal of Environmental Quality* 40(2): 647-652.
- Ravichandran, M. (2004), Interactions between mercury and dissolved organic matter: a review, *Chemosphere* 55(3): 319-331.
- Reddy, M. M. and Aiken, G. R. (2001), Fulvic Acid-Sulfide Ion Competition for Mercury Ion Binding in the Florida Everglades, *Water, Air, & Soil Pollution* 132(1): 89-104.
- Remy, S., Prudent, P. and Probst, J.-L. (2006), Mercury speciation in soils of the industrialised Thur River catchment (Alsace, France), *Applied Geochemistry* 21(11): 1855-1867.
- Renneberg, A. J. and Dudas, M. J. (2001), Transformations of elemental mercury to inorganic and organic forms in mercury and hydrocarbon co-contaminated soils, *Chemosphere* 45(6-7): 1103-1109.
- Rinklebe, J., During, A., Overesch, M., Du Laing, G., Wennrich, R., Stärk, H.-J. and Mothes, S. (2010), Dynamics of mercury fluxes and their controlling factors in large Hg-polluted floodplain areas, *Environmental Pollution* 158(1): 308-318.
- Ruiz, O. N. and Daniell, H. (2009), Genetic engineering to enhance mercury phytoremediation, *Current Opinion in Biotechnology* 20(2): 213-219.
- Saeid, A. (2004), Kinetic models of sorption: a theoretical analysis, *Journal of Colloid and Interface Science* 276(1): 47-52.
- Santoro, A., Terzano, R., Blo, G., Fiore, S., Mangold, S. and Ruggiero, P. (2010), Mercury speciation in the colloidal fraction of a soil polluted by a chlor-alkali plant: a case study in the South of Italy, *Journal of Synchrotron Radiation* 17(2): 187-192.
- Santoro, A., Terzano, R., Medici, L., Beciani, M., Pagnoni, A. and Blo, G. (2012), Colloidal mercury (Hg) distribution in soil samples by sedimentation field-flow fractionation coupled to mercury cold vapour generation atomic absorption spectroscopy, *Journal of Environmental Monitoring* 14(1).
- Sarkar, D., Essington, M. E. and Misra, K. C. (2000), Adsorption of Mercury(II) by Kaolinite, *Soil Sci. Soc. Am. J.* 64(6): 1968-1975.
- Schlüter, K. (2000), Review: evaporation of mercury from soils. An integration and synthesis of current knowledge, *Environmental Geology* 39(3): 249-271.
- Schroeder, W. H. and Munthe, J. (1998), Atmospheric mercury-An overview, *Atmospheric Environment* 32(5): 809-822.
- Schroeder, W. H., Munthe, J. and Lindqvist, O. (1989), Cycling of mercury between water, air, and soil compartments of the environment, *Water, Air, & Soil Pollution* 48(3): 337-347.

- Schuster, E. (1991), The behavior of mercury in the soil with special emphasis on complexation and adsorption processes - A review of the literature, *Water, Air, & Soil Pollution* 56: 667-680.
- Senevirathna, W. U., Zhang, H. and Gu, B. (2011), Effect of carboxylic and thiol ligands (oxalate, cysteine) on the kinetics of desorption of Hg(II) from kaolinite, *Water, Air, & Soil Pollution* 215(1): 573-584.
- Shaw, S. A., Al, T. A. and MacQuarrie, K. T. B. (2006), Mercury mobility in unsaturated gold mine tailings, Murray Brook mine, New Brunswick, Canada, *Applied Geochemistry* 21(11): 1986-1998.
- Skylberg, U. (2010), Mercury Biogeochemistry in Soils and Sediments, *Developments in Soil Science* 34.
- Skylberg, U. (2012), *Chemical Speciation of Mercury in Soil and Sediment*. Environmental Chemistry and Toxicology of Mercury, John Wiley & Sons, Inc.: 219-258.
- Skylberg, U., Qian, J., Frech, W., Xia, K. and Bleam, W. F. (2003), Distribution of mercury, methyl mercury and organic sulphur species in soil, soil solution and stream of a boreal forest catchment, *Biogeochemistry* 64(1): 53-76.
- Slowey, A. J., Johnson, S. B., Rytuba, J. J. and Brown, G. E. (2005a), Role of Organic Acids in Promoting Colloidal Transport of Mercury from Mine Tailings, *Environmental Science & Technology* 39(20): 7869-7874.
- Slowey, A. J., Rytuba, J. J. and Brown, G. E. (2005b), Speciation of Mercury and Mode of Transport from Placer Gold Mine Tailings, *Environmental Science & Technology* 39(6): 1547-1554.
- Song, X. and Van Heyst, B. (2005), Volatilization of mercury from soils in response to simulated precipitation, *Atmospheric Environment* 39(39): 7494-7505.
- Sorkhoh, N. A. (2010), Phytoremediation of mercury in pristine and crude oil contaminated soils: Contributions of rhizobacteria and their host plants to mercury removal, *Ecotoxicology and environmental safety* 73(8): 1998.
- Subir, M., Ariya, P. A. and Dastoor, A. P. (2011), A review of uncertainties in atmospheric modeling of mercury chemistry I. Uncertainties in existing kinetic parameters – Fundamental limitations and the importance of heterogeneous chemistry, *Atmospheric Environment* 45(32): 5664-5676.
- Sunderland, E. M. (2009), Mercury sources, distribution, and bioavailability in the North Pacific Ocean: Insights from data and models, *Global biogeochemical cycles* 23(2).
- Terzano, R., Santoro, A., Spagnuolo, M., Vekemans, B., Medici, L., Janssens, K., Göttlicher, J., Denecke, M. A., Mangold, S. and Ruggiero, P. (2010), Solving mercury (Hg) speciation in soil samples by synchrotron X-ray microspectroscopic techniques, *Environmental Pollution* 158(8): 2702-2709.
- Tsakiroglou, C. D. and Payatakes, A. C. (1998), Mercury intrusion and retraction in model porous media, *Advances in Colloid and Interface Science* 75(3): 215-253.
- Ullrich, S. M., Tanton, T. W. and Abdrashitova, S. A. (2001), Mercury in the Aquatic Environment: A Review of Factors Affecting Methylation, *Critical Reviews in Environmental Science and Technology* 31(3): 241-293.
- UNEP (2002), Global Mercury Assessment. Geneva, Switzerland.
- Van Geel, P. J. and Roy, S. D. (2002), A proposed model to include a residual NAPL saturation in a hysteretic capillary pressure-saturation relationship, *Journal of Contaminant Hydrology* 58(1-2): 79.
- Walvoord, M. A., Andraski, B. J., Krabbenhoft, D. P. and Striegl, R. G. (2008), Transport of elemental mercury in the unsaturated zone from a waste disposal site in an arid region, *Applied Geochemistry* 23(3): 572-583.

- Waples, J. S., Nagy, K. L., Aiken, G. R. and Ryan, J. N. (2005), Dissolution of cinnabar (HgS) in the presence of natural organic matter, *Geochimica et Cosmochimica Acta* 69(6): 1575-1588.
- Weaver, J. W., Charbeneau, R. J., Tauxe, J. D., Lien, B. K. and Provost, J. B. (1994), *The Hydrocarbon Spill Screening Model (HSSM). Volume 1: User's Guide*, Robert S. Kerr Environmental Research Laboratory, Office of Research and Development, U.S. Environmental Protection AgencyAda, Oklahoma 74820, p.
- Wipfler, E. L. and van der Zee, S. E. A. T. M. (2001), A set of constitutive relationships accounting for residual NAPL in the unsaturated zone, *Journal of Contaminant Hydrology* 50(1-2): 53.
- Xia, K., Skyllberg, U. L., Bleam, W. F., Bloom, P. R., Nater, E. A. and Helmke, P. A. (1999), X-ray Absorption Spectroscopic Evidence for the Complexation of Hg(II) by Reduced Sulfur in Soil Humic Substances, *Environmental Science & Technology* 33(2): 257-261.
- Yin, Y., Allen, H. E., Huang, C. P. and Sanders, P. F. (1997a), Adsorption/Desorption Isotherms of Hg(II) by Soil, *Soil Science* 162(1): 35-45.
- Yin, Y., Allen, H. E., Huang, C. P., Sparks, D. L. and Sanders, P. F. (1997b), Kinetics of Mercury(II) Adsorption and Desorption on Soil, *Environmental Science & Technology* 31(2): 496-503.
- Zhang, H. and Selim, H. M. (2006), Modeling the Transport and Retention of Arsenic (V) in Soils, *Soil Sci. Soc. Am. J.* 70(5): 1677-1687.
- Zhang, L., Wright, L. P. and Blanchard, P. (2009), A review of current knowledge concerning dry deposition of atmospheric mercury, *Atmospheric Environment* 43(37): 5853-5864.

## 6 Annex

### 6.1 NAPL migration and entrapment

There is no major difference in behaviour between LNAPL and DNAPL in the vadose zone. Modeling NAPL migration usually involves complex multi-phase models, in which relative permeabilities, saturations and pressures determine NAPL movement.

Miller (1998) provided a review of multiphase flow and transport models (of which most were used for studying organic compounds). The standard approach relies on continuum balance equations about a representative elementary volume, empirical equations of state and constitutive relations (pressure-saturation-permeability). In the context of modeling NAPL as a source of contamination in the unsaturated zone, it is important that the constitutive relations account for residual (and entrapped) NAPL<sup>1</sup>. Wipfler and van der Zeze (2001) defined a critical NAPL pressure at the transition between free NAPL and residual NAPL; i.e. all NAPL becomes residual below a critical total liquid saturation. However, Oostrom et al. (2005) found no experimental evidence of such a critical total liquid saturation. Oostrom et al. (2005) found a good agreement with experimental observations using the models of Van Geel and Roy (2002) and of Lenhard et al. (2004). These models stem from an existing hysteretic relative permeability-saturation-capillary pressure model and assume the formation of residual NAPL during NAPL drainage and imbibition, respectively.

Very few studies are specific to mercury for NAPL characterization and modeling. A notable exception is the work of Devasena and Nambi (2010). They suggested that mercury migration was influenced only by gravity and capillary forces and not by viscous forces. Following their experiment, the capillary forces dominate and lead to mercury entrapment. They measured Hg residual saturation of 0.04 and 0.08 for coarse and fine sand, respectively. They also estimated capillary forces to be  $1.22 \times 10^4$  times higher than gravity force for mercury NAPL in saturated soil media (coarse sand). The immobile mercury can then be a persistent source of contamination. However, these findings were obtained in saturated media. Mercer and Cohen (1990) noted that in general, residual saturation in the vadose zone is smaller than in the saturated zone. Moreover residual saturation in the vadose zone increases with decreasing intrinsic permeability, effective porosity and moisture content (Mercer and Cohen, 1990).

Mercury was suggested to produce lower residual NAPL compared to other DNAPLs (e.g. PCE, TCE) (Devasena and Nambi, 2010). Larger Bond number (Bond number relates gravity and capillary forces) for a mercury-water system than for PCE-water systems reflect larger density differences, which lead to less residual saturation.

- 1) Nambi [AGU abstract] *“Mercury-water systems exhibited a low residual saturation of 0.02 and 0.07 as compared to 0.16 and 0.27 for PCE-water systems. Less residual mercury saturation, lack of apparent hysteresis in capillary pressure saturation curves and large variation in van Genuchten’s parameters ‘ $\alpha$ ’ (inverse of displacement pressure) and ‘ $n$ ’ (pore size distribution index) for mercury-water systems compared to PCE-water*

---

<sup>1</sup> Lenhard et al. (2004) make the distinction between residual, entrapped and free NAPL. Residual NAPL is immobile NAPL in small pores and pore wedges (i.e. can be continuous), formed during imbibition. Entrapped NAPL is immobile NAPL occluded by water (i.e. discontinuous). Free NAPL is mobile NAPL, moving in larger pores.

*systems were observed. These anomalies between the two systems elucidate that the capillary trapping is equally dependent on the fluid characteristics especially for high density immiscible fluids. Gravity force nevertheless a predominant controlling factor in the migration of highly dense mercury, is counteracted by not less trivial capillary force which was  $1.22 \times 10^4$  times higher than gravity force. The capillary forces thus surmount the gravity forces and cause entrapment of mercury in the soil pores even in homogeneous porous medium system. Bond number (Bond number relates gravity and capillary forces) for mercury-water system was found to 2.5 times higher than PCE-water systems. Large density differences between mercury and water lead to high Bond number and thus less residual saturation. Capillary number (Capillary number relates viscous and capillary forces) was found to be less for mercury-water systems. Literature review unveils that low Capillary number does not influence non wetting residual saturation. But for high density mercury with natural infiltration, even low Capillary number influences residual saturation. With the alarming increase in number of mercury spill sites, results of this study showed a better understanding of the capillary entrapment phenomena and the extent of influence of each predominant force during displacement of highly dense mercury. The fundamental inputs to NAPL entrapment models were generated in this study for mercury for the first time.”*

It may be interesting to have an idea of the travel time needed for Hg NAPL to migrate in the subsurface. Gerhard et al. (2007) numerically studied DNAPL mobility in a sandy aquifer. They simulated DNAPL migration in aquifer for the following compounds: TCE, PCE, 1,1 DCE, chlorobenzene, PCB and coal tar. DNAPL density and viscosity were identified as parameters having a significant influence on time scales of migration. Gerhard et al. (2007) showed that DNAPL migration (infiltration rate in saturated zone, in  $\text{m}^3 \text{y}^{-1}$ ) could be plotted as a function of DNAPL mobility  $M_D$  ( $\text{s m}^{-3}$ ), defined as  $M_D = \frac{\Delta\rho}{\mu_D}$ , where  $\Delta\rho$  is the difference between DNAPL and water densities ( $\text{kg m}^{-3}$ ) and  $\mu_D$  is DNAPL viscosity ( $\text{Pa s}$ ). If the properties of mercury ( $\rho_{\text{Hg}} = 13546 \text{ kg m}^{-3}$  at  $20^\circ\text{C}$  and  $\mu_{\text{Hg}} = 1.526 \times 10^{-3} \text{ Pa s}$ ;  $M_D = 8.24 \times 10^6 \text{ s m}^{-3}$ ) are compared to those of the DNAPLs analysed by Gerhard et al. (2007), mercury would be classified as a highly mobile compound; i.e. having relatively short time scales for “subsurface invasion”. However, this extrapolation has to be considered with caution because the behaviour of Hg NAPL in the subsurface may not be directly comparable to other DNAPLs, due to the unique properties of mercury<sup>2</sup> (Devasena and Nambi, 2010).

Given that the parameterisation of permeability-saturation-capillary relations seems problematic for Hg NAPLs, one could use simpler models of NAPL migration in the vadose zone. For example, the KOPT module of the HSSM model (Weaver et al., 1994) can simulate the migration of a NAPL front in the vadose zone. To investigate the rate of downward migration of a Hg NAPL in a soil of texture “light sand loam”, HSSM was run with the parameterization presented in Table 1. Figure 4 shows the NAPL migration front at different time steps. It can be seen that, starting from a depth of 0.5 m, the front migration reaches 1 m after 2 days; and about 3.3 m (i.e. a 2.8 m downward migration) after 1 year. The downward migration rate is of course highly sensitive to the NAPL release conditions. In a simulation with a 0.1 m ponding depth, it takes ~40 days for Hg NAPL to migrate 0.5 m downwards, and the migration front is ~0.77 m deep after 1 year (i.e. 0.67 m migration). In a simulation with 1 m ponding depth, it takes only ~30 days for the NAPL front to migrate 2.8 m downward (results not shown).

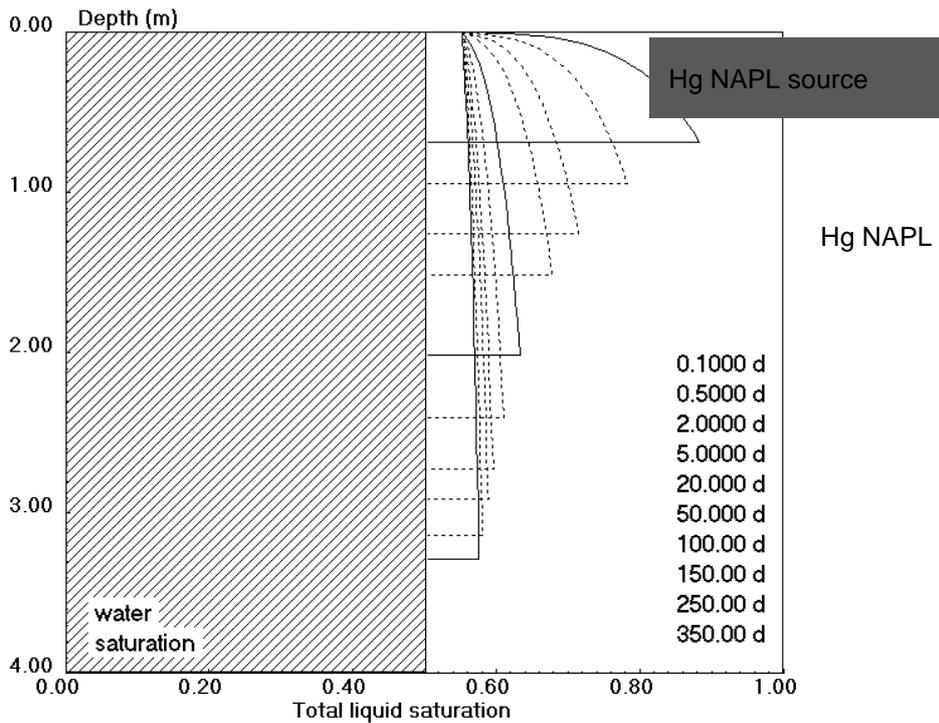
Similar simulations were performed by Marsman et al. (2012), who tested different infiltration scenarios of Hg DNAPL using the model STOMP.

---

<sup>2</sup> For example, mercury is the only NAPL which does not preferentially wet solid particles.

**Table 1 – Parameters of the HSSM model for simulating a test case of Hg NAPL migration in the vadose zone.**

Parameter	Unit	Value	Comment
Water infiltration	m/d	0.001096	Equivalent to 400 mm/y
$\theta_r$	-	0.034	Soil parameters derived for a light sand loam
van Genuchten $\alpha$	1/m	2.21	
van Genuchten $n$	-	1.423	
$K_{sat}$	m/d	0.49	
Max. relative permeability	-	0.5	Default; ensures that the soil is never saturated
Hg NAPL density	g/cm <sup>3</sup>	13.546	At 20°C (Ebadian et al., 2001)
Hg dynamic viscosity	cP	1.554	At 20°C (Ebadian et al., 2001)
Hg NAPL solubility	mg/L	0.0662	(Clever, 1985)
Hg NAPL surface tension	dyn/cm	436	At 20°C (Ebadian et al., 2001)
Hg NAPL residual saturation in vadose zone	-	0.05	Chosen lower than values for saturated zone in (Devasena and Nambi, 2010); accounting for lower residual saturation in vadose zone than in aquifer (Mercer and Cohen, 1990)
NAPL ponding depth	m	0.5	NAPL release is simulated as a 0.5 m "landfill" at the soil surface
Lower depth of NAPL zone	m	0.5	



**Figure 4 – Hg NAPL saturation profile at different time steps, for an illustrative simulation with parameters given in Table 1.**

In practical applications, if Hg contamination in the vadose zone can be assumed to be historical (i.e. only immobile Hg is left), NAPL residual saturation is the main parameter to estimate. Residual saturation can then be used as input to parameterize Hg release to the aqueous and gas phases.

